



## Review

# HIV in the CNS: Pathogenic relationships to systemic HIV disease and other CNS diseases

Dianne M Rausch<sup>1</sup> and Miriam R Davis<sup>2</sup>

<sup>1</sup>National Institute of Mental Health, Center for Mental Health Research on AIDS, Bethesda, Maryland, USA; and <sup>2</sup>Department of Epidemiology and Biostatistics, School of Public Health and Health Services, George Washington University, Washington, DC, USA

**Research on the pathogenesis of the human immunodeficiency virus (HIV) infection of the central nervous system (CNS) has reached a pivotal stage. While the incidence of HIV dementia appears to be declining, the prevalence of milder, yet debilitating, neuropsychological impairments may rise as individuals infected with HIV live longer. There are also concerns about CNS reservoirs of latently infected cells. Building upon progress in understanding HIV neuropathogenesis, the time is ideal to expand research on the interrelationships between the CNS and systemic HIV disease, and extend the boundaries of this research to the neuropathogenic similarities between HIV and other CNS inflammatory diseases. Neuropathogenic insights gained from these pursuits can spawn new treatment strategies for HIV/CNS disease as well as potentially other diseases of the nervous system.** *Journal of NeuroVirology* (2001) 7, 85–96.

**Keywords:** HIV; dementia; Alzheimer's disease; multiple sclerosis; trafficking; chemokines

## Introduction

HIV infection of the CNS produces a range of cognitive, motor, and behavioral abnormalities (Navia *et al*, 1986a; Price *et al*, 1988). HIV-associated dementia (HAD) is the most severe manifestation, typically found in late stages of AIDS. But during the course of HIV disease—even early after infection—more subtle neuropsychological (NP) impairments are detectable (Grant *et al*, 1995). NP impairments can be disabling (Heaton *et al*, 1994; Albert *et al*, 1995; Heaton *et al*, 1996), can interfere with patient adherence to complex medication regimens, and are an independent risk factor for mortality (Ellis *et al*, 1997).

The advent of highly active antiretroviral therapy (HAART) in 1995 has led to striking reductions in plasma viral load, opportunistic infections, and mor-

tality from AIDS (Montaner *et al*, 1998; CDC, 1999; Powderly, 2000). During this era there also appears to be a reduction in the incidence of HAD (Dore *et al*, 1999; Sacktor *et al*, 1999b). However, as patients live longer, there is reason to predict growing prevalence of NP impairment and/or HAD. Further, because of poor drug penetration of the blood-brain barrier (BBB) or poor accumulation within the CNS, concerns persist about the possible evolution of drug-resistant virus in the CNS (Richman, 1996; Kepler and Perelson, 1998; Schrager and D'Souza, 1998). There are currently no FDA-approved treatments expressly designed for the CNS, although several nonantiretroviral drugs are in clinical trials.

The refinement of *in vitro* and animal models, the identification of rapidly increasing roles for chemokines and their receptors, and improvements in viral load monitoring have ushered in significant advances in our understanding of HIV neuropathogenesis. To build on that knowledge, researchers are poised to look beyond the confines of the CNS. New sets of questions can be posed about potential dynamic interrelationships between the CNS and the periphery. Researchers also can branch out to focus on common pathogenic threads uniting HIV neuropathogenesis with that of other neurological

---

This article is not subject to U.S. copyright laws.  
Address correspondence to DM Rausch, National Institute of Mental Health, Center for Mental Health Research on AIDS, 6100 Executive Blvd, Room 6212, MSC 9623, Bethesda, MD 20892-9623, USA. E-mail: dr89b@nih.gov  
Received 17 October 2000; revised 11 December 2000; accepted 19 December 2000

disorders, most notably multiple sclerosis and Alzheimer's disease. The pathogenic similarities between seemingly disparate disorders, with remarkably different etiologies, can inspire new hypotheses about HIV neuropathogenesis and its treatment.

## HIV in the CNS

HIV commonly invades the CNS (Wiley *et al*, 1999; Masliah *et al*, 2000), and it does so early after peripheral infection (see reviews by Griffin, 1998; Kolson *et al*, 1998). HIV has been detected in the brain as early as 15 days after accidental intravenous inoculation (Davis *et al*, 1992). Early penetration into the CNS also holds true following peripheral inoculation of rhesus macaques with SIV (Hurtrel *et al*, 1991; Lackner *et al*, 1991; Sharer *et al*, 1991). The chief cellular targets of HIV infection within the CNS are microglia/macrophages (MG/MP) (Kolson *et al*, 1998). MG/MP is a generic term for monocyte-derived cells that, upon their migration into the CNS, differentiate into resident microglia, perivascular microglia and macrophages, meningeal macrophages, and choroid plexus macrophages, among others (Hickey, 1999a).

### *Indirect mechanisms of neuropathogenesis*

More than a decade ago, it became clear that the neuron dysfunction or death that underlies clinical symptoms of HIV/CNS disease cannot result from direct infection of neurons. The reigning model of HIV neuropathogenesis attributes neuron dysfunction or death to the *indirect* consequences of infection of MG/MP (Giulian *et al*, 1990; Pulliam *et al*, 1991). Under this model, MG/MP release a barrage of cytokines and other soluble factors, including HIV proteins, that, in high concentrations and over extended periods, are toxic to nearby neurons. Cytokines and other inflammatory factors are also produced and released by *uninfected* MG/MP that are activated by cytokines or by soluble HIV proteins (Nuovo and Alfieri, 1996; Yeh *et al*, 2000).

The *combined* influence of both HIV infection and activation of immune-competent cells explains the important finding that HIV infection alone cannot fully account for the degree of dementia (Glass *et al*, 1995). In this autopsy study, the presence of activated MG/MP was better correlated with the degree of dementia than was viral load. HIV infection, in other words, sets the stage for secretion of inflammatory products, which in some cases leads to neuronal apoptosis, by either *infected* MG/MP (Pulliam *et al*, 1991), or by *uninfected*, yet immune-activated, MG/MP (Yeh *et al*, 2000). The number of activated cells is vastly increased by either astrocytosis or by influx of leukocytes (see next section).

The repertoire of candidate neurotoxins released by immunocompetent cells is large. Neurotoxic HIV proteins include gp120 envelope glycoprotein, Tat, and Nef (Kolson *et al*, 1998). The cytokines and

other soluble factors that are significantly elevated in the brains or CSF of HIV-infected individuals include pro-inflammatory cytokines (TNF- $\alpha$  and IL1- $\beta$ ), chemokines (see next section), arachidonic acid, platelet activating factor, quinolinic acid, and nitric oxide (see reviews Kolson *et al*, 1998; Zheng and Gendelman, 1997). Quinolinic acid levels correlate with severity of impairments or dementia in humans (Sei *et al*, 1995) and motor impairment in animals (Heyes *et al*, 1992; Rausch *et al*, 1994). Release of TNF- $\alpha$  by MG/MP also may be critical, as levels of TNF- $\alpha$  mRNA at autopsy correlate with the severity of dementia (Wesselingh *et al*, 1993). Further, TNF- $\alpha$  may increase white matter pallor seen at autopsy because, on the basis of *in vitro* studies, its release from HIV-infected MG/MP kills oligodendrocytes (Wilt *et al*, 1995).

### *Chemokines*

One of the prominent neuropathological findings with HIV infection is perivascular infiltration of monocytes across the BBB (Price *et al*, 1988). Recent research has highlighted the role of chemokines as key regulators of monocyte recruitment across the BBB into the CNS. This chemoattractant role for chemokines is distinct from their numerous other roles, most notably as coreceptors (with CD4) for entry of HIV into monocytes and lymphocytes. During infection of the CNS, release of specific chemokines induces selective migration across the BBB by those leukocytes with corresponding chemokine receptors. Chemokines and their receptors do not work alone, as the selectins and integrins (and their receptors) also participate in a multistep process of immune cell infiltration (Luster, 1998).

$\beta$ -Chemokines figure prominently in HIV infection of the CNS (Meucci *et al*, 1998; Hesselgesser and Horuk, 1999). They are selective attractants for monocytes and lymphocytes. The CNS cells expressing  $\beta$ -chemokines (or their cognate receptors) are neurons, MG/MP, and the cells forming the BBB (endothelial cells and astrocytes) (Hesselgesser and Horuk, 1999).

The expression of several chemokines is upregulated during HIV infection, and this enhanced expression correlates with dementia. A groundbreaking autopsy study found the brains of HIV patients with dementia to have elevated levels of MIP-1 $\alpha$  and MIP-1 $\beta$ , compared with patients without dementia (Schmidtmayerova *et al*, 1996). In the same study cultured monocytes infected with HIV produced elevated levels of the same  $\beta$ -chemokines. Similarly, studies in the SIV model found MIP-1 $\alpha$ , MIP-1 $\beta$ , RANTES, MCP-3 and the  $\alpha$ -chemokine IP-10 in brain (Sasseville *et al*, 1996; Westmoreland *et al*, 1998). IP-10 is one of the  $\alpha$ -chemokines that selectively attracts activated T lymphocytes (Luster, 1998). IP-10 was found to be elevated in the CSF of HIV-infected patients with neurological deficits

(Kolb *et al*, 1999). Other CSF studies of HIV patients have found elevated concentrations of  $\beta$ -chemokines. Higher CSF levels of either MCP-1 or RANTES, or both, were found to be correlated with the degree of encephalitis (Cinque *et al*, 1998) and dementia (Conant *et al*, 1998; Kelder *et al*, 1998). MCP-1, MIP-1 $\alpha$ , and RANTES were localized by immunocytochemistry to be most abundant in and around microglial nodules, a histopathological sign of HIV infection at autopsy (Sanders *et al*, 1998).

### Pathogenic relationships between CNS and systemic HIV disease

For years, research on HIV-associated dementia (HAD) naturally focused on events within the CNS. It also focused primarily on pathogenic mechanisms occurring either early or late in HIV disease. But recent research on the existence of systemic HIV reservoirs (Chun *et al*, 1997; Finzi *et al*, 1997; Wong *et al*, 1997b) and leukocyte trafficking through the CNS has begun to spur interest in the possibility of dynamic interrelationships between the CNS and systemic disease. Such interrelationships, in fact, may be occurring throughout the course of HIV disease, not just early after infection or at end stages. Interrelationships—especially during asymptomatic phases of HIV disease—are vital to study because, with treatment advances, the burden of the epidemic in the US has shifted to a more chronic course (Rausch and Stover, 2000).

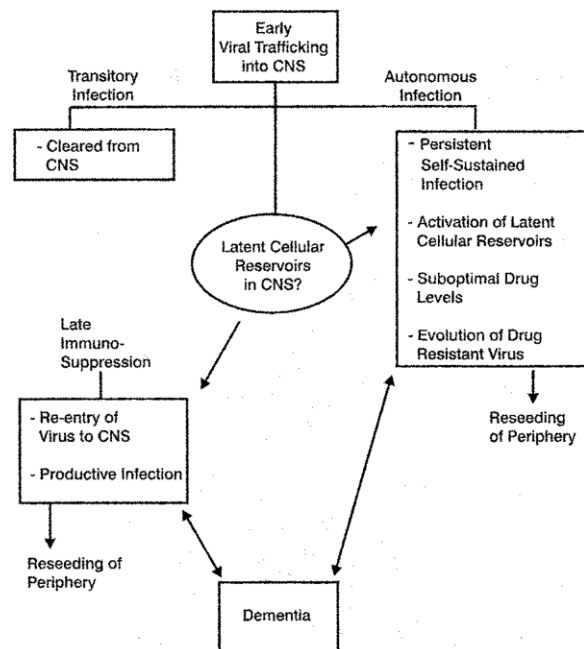
CNS trafficking refers to the movement of immune cells (or virus) from peripheral blood, across the BBB, through brain parenchyma, and then back into the periphery. Until the last decade, the dogma was that the normal CNS was immunologically privileged and that immune cells did not traffic into and out of the brain (Miller, 1999). Now there is greater recognition of immune cell trafficking as a dynamic process involving numerous cell types with divergent functions, cell surface markers, migratory capacity, CNS location, and turnover kinetics (Hickey, 1999a). Our understanding of the details of monocyte and other immune cell trafficking, under normal physiology and under the influence of HIV infection, is still rudimentary. It is not established, for instance, whether the virus remains in the CNS or is cleared from the brain (Zink *et al*, 1998). The most formidable problem for research is that access to human CNS tissue, except at autopsy, is severely limited. A further limitation is that CSF has uncertain value for studying events within the CNS, especially prior to advanced HIV disease (Price and Staprans, 1997; Ellis *et al*, 2000). Many of the questions that follow can be addressed in the SIV animal model.

#### Trafficking and autonomous versus transitory infection

HIV enters the CNS early (see prior description), before systemic infection typically is recognized and

treatment initiated. Yet, HAD may not develop until years thereafter, when there is substantial viral replication in the CNS (Wiley *et al*, 1998; Wiley *et al*, 1999). After its early entry, what happens to the virus in the CNS, and what is its relationship to the periphery over the natural history of the disease? Does the virus entering early account for later CNS manifestations? This would be an *autonomous* infection, i.e., one that is persistent and self-sustaining, not dependent on subsequent trafficking of virus from the systemic circulation. Or is initial CNS viral infection short-lived, readily cleared from the CNS, and later dependent on continuous or repeated trafficking from the periphery to sustain CNS infection giving rise to HAD? This type of CNS infection would be *transitory* (see Figure 1).

The conceptual dichotomy between autonomous and transitory infection was first articulated by Price and Staprans (1997). As with any dichotomy, the answer may not exclusively be one or the other, but some combination of both. Understanding the extent of autonomous versus transitory infection in relation to disease stage has important implications for treatment and eradication of HIV. If autonomous infection in the CNS is the major culprit in HAD or milder forms of impairment, then treatment strategies should emphasize new medications directly for the CNS. If, on the other hand, virus is removed early and without clinical significance, but later trafficking of virus into the CNS is the culprit in HAD, then more effective control of HIV in the periphery is warranted. As long as peripheral control over viral replication is achieved, according to this line of reasoning, HAD would not develop (Gartner, 2000).



**Figure 1** Transitory versus autonomous infection after HIV trafficking into the CNS.

To disentangle the relative roles of autonomous and transitory infection over the course of HIV disease would require repeated CNS and peripheral tissue sampling at different times during infection. CNS sampling can be accomplished in experimental animal models but it is not possible in humans. Thus evidence in humans favoring one or other type of infection at this point is largely indirect.

Autonomous infection of the CNS is thought to be plausible because of the cell types infected, their turnover characteristics, and because of the protected environment of the CNS. The most commonly infected cells (monocyte derivatives) are not lysed during production of viral progeny. They are long-lived, with some cell types (e.g., parenchymal microglia) persisting possibly for decades (Hickey *et al*, 1992; Lassman *et al*, 1993). Further, the CNS is to some extent both immunologically and pharmacologically protected from the systemic circulation by the BBB (Miller, 1999). These features generate concerns that, especially in a CNS environment of sub-optimal drug levels, an autonomous infection could proceed unchecked, leading to evolution of drug-resistant viral mutations (Richman, 1996; Kepler and Perelson, 1998; Ellis *et al*, 2000). Such mutations would threaten *both* the CNS and the periphery should mutated virus traffic out of the CNS. Reseeding from the CNS to the periphery late in disease has been demonstrated with the SIV model (Zink *et al*, 1997).

Compartmentalization of HIV in the CNS has been found in numerous studies showing certain viral sequences, including drug resistance codons, to be different from those in other compartments (Pang *et al*, 1991; Korber *et al*, 1994; Wong *et al*, 1997a; Hughes *et al*, 1997; van't Wout *et al*, 1998; Liu *et al*, 2000). There are also reported genetic differences across distinct regions of the brain (Morris *et al*, 1999; Shapshak *et al*, 1999). The limitation of many of these studies to understanding the extent of transitory versus autonomous infection is that they were done on a small number of individuals, generally with advanced disease at autopsy. Further, most were cross-sectional, so it is difficult to address the timing of viral entry into the CNS. Longitudinal studies—with advanced imaging and repeated blood and CSF sampling—are clearly warranted to determine whether and to what extent productive replication is occurring throughout disease, and what is the source of virus—internal to the CNS or from blood trafficking? What sites, like the choroid plexus (Petito *et al*, 1999), might serve as a reservoir for CNS infection?

It is also vital to examine viral and host factors—throughout disease—that affect HIV and immune cell trafficking between the systemic circulation and the CNS. Viral strain (Zink *et al*, 1998), viral load, and integrity of the BBB (Petito and Cash, 1992; Dallasta *et al*, 1999) are likely important, as is the activation state of monocytes. Monocyte migration across an artificial BBB, for example, was shown to in-

crease by 20-fold when monocytes were activated (Persidsky *et al*, 1997). Monocyte activation in this study was even more important than viral infection in enhancing monocyte trafficking. The activation state of monocytes affects the profile of cytokines that they produce and secrete (Frankenberger *et al*, 1996) and, as noted earlier, their neurotoxicity.

A related question is whether there is a reservoir of latently infected cells (monocytes, T cells, or astrocytes) within the CNS. If so, can latently infected cells be activated to sustain autonomous infection? Latent infection, a means for HIV to evade host defenses, refers to proviral DNA incorporated into the host cell's genome in a transcriptionally silent form until later activation and replication (Chun and Fauci, 1999). Peripheral tissues, such as lymph nodes, bone marrow and testis, hold latent viral reservoirs within resting CD4+ T cells. Because of T cells' slow turnover (months to years), latent infection may be responsible for lifelong persistence of HIV, despite HAART's suppression of virus in actively replicating cells (Finzi *et al*, 1999).

Strong evidence for peripheral cellular reservoirs has clearly raised the specter of CNS cellular reservoirs (Schrager and D'Souza, 1998; Chun *et al*, 2000). Yet, the precise nature and extent of CNS reservoirs are elusive. There appears to be a small proviral load of HIV in the brain in *asymptomatic* HIV disease (Bell *et al*, 1993; Donaldson *et al*, 1994; Sinclair *et al*, 1994). The question is whether and under what circumstances it is sufficient to rekindle productive CNS infection and its clinical manifestations. In the absence of certainty, even the possibility of CNS reservoirs warrants the development of explicit CNS treatment strategies aimed at their reactivation and eradication, similar to those being tested for the periphery, e.g., IL-2 coupled with anti-retroviral therapy (Chun *et al*, 1999).

#### *Impact of HAART*

The introduction of HAART, as noted earlier, appears to have coincided with a reduction in the incidence of HAD. Clinical trials have found HAART to alleviate some NP impairments (Ferrando *et al*, 1998; Tozzi *et al*, 1999; Sacktor *et al*, 2000), although it is not clear if protease inhibitor in the combination confers any distinct advantages over combination therapy without protease inhibitor (Sacktor *et al*, 1999a). HAART's impact on the full array CNS impairments has not yet been studied.

HAART's apparent benefits are perplexing in light of poor CNS penetration of many of its constituent drugs (Enting *et al*, 1998; Flexner, 1998; Aweeka *et al*, 1999). Zidovudine (ZVD) and indinavir appear to have among the best CNS penetration, but both are substrates for active efflux (Banks, 1999; Martin *et al*, 1999). Their suppression of CNS viral replication cannot be studied in patients during asymptomatic stages. It is thus unclear whether HAART's seeming benefits may stem from partial, albeit sufficient,

control over viral replication in the CNS or from control of viral replication in peripheral compartments (thereby curtailing trafficking into the CNS). Another area to investigate is HAART's impact on peripheral monocyte activation and cytokine profiles. There is suggestive evidence that HAART shifts the cytokine profile of monocytes and T cells (Imami *et al*, 1999). Key areas of investigation are HAART's impact on monocytes, including their cytokine profiles, capacity to traffic into the CNS, and their neurotoxicity.

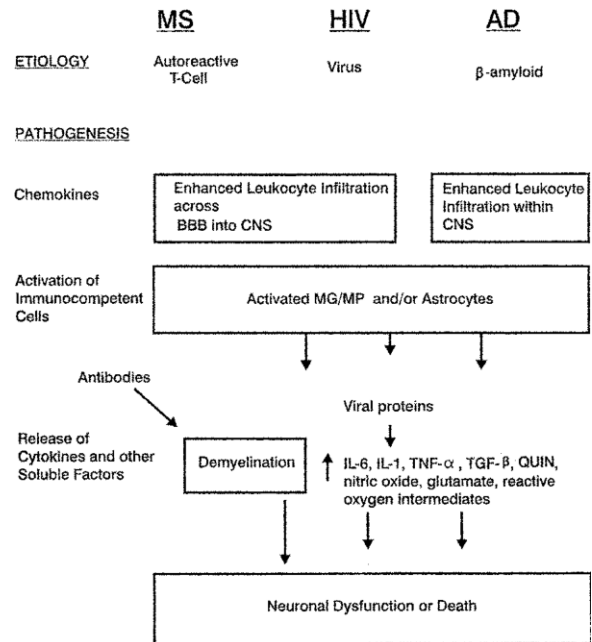
In summary, many questions for research surround trafficking of HIV into the CNS and the interrelationships with the periphery over the course of HIV disease. The impact of HAART on trafficking and on the nature and extent of CNS manifestations is also key. Better understanding of neuropathogenesis in the era of HAART is crucial as the nature of the HIV epidemic shifts to a more long-term course.

### Pathogenic relationships between HIV and other CNS diseases

Until recently neuroscientists tended to emphasize the neuropathogenic differences across distinct CNS diseases. Attention was understandably drawn to the differences in an effort to characterize in detail neuropathogenic mechanisms unique to each. Now, a change in perception is emerging, building on momentum from advances in immunology, molecular biology, and neuroscience. Researchers have begun to recognize, and capitalize upon, some intriguing parallels in the neuropathogenesis of disparate CNS diseases (Dickson *et al*, 1993; Mrak and Griffin, 1997; Cotter *et al*, 1999; Gonzalez-Scarano and Baltuch, 1999; Hesselgesser and Horuk, 1999).

This section describes some of the key similarities in neuropathogenesis between HIV disease and two other neurodegenerative diseases, multiple sclerosis (MS) and Alzheimer's disease (AD). The section highlights the overlaps in three critical areas: chemokines and leukocyte chemotaxis; activation of inflammatory cells and release of soluble cytotoxins; and mechanisms of neuron dysfunction and death (Figure 2). The choice of MS and AD is for illustrative purposes, for these areas of overlap may well extend to other neurological disorders.

Enthusiasm surrounding pathogenic similarities should not lose sight of fundamental differences between HIV, MS, and AD. The foremost difference rests with etiology. The etiological agent in HIV disease is the virus; with MS, the etiology is not established, but evidence favors the early role of autoreactive T lymphocytes (CD4+) directed at myelin antigens (Williams *et al*, 1994; Brosnan and Raine, 1996; Conlon *et al*, 1999; Lassmann, 1999). With AD, the etiology is also not fully known, but most evidence points to genetic mutations which lead to the production of extracellular aggregates of  $\beta$ -amyloid protein (Selkoe,



**Figure 2** Etiology and neuropathogenesis of MS, HIV, and AD. Despite different etiologies, there are pathogenic similarities.

1999). The three diseases also exhibit differences in symptomatology, diagnosis, and clinical course, although there are some overlaps in the nature of cognitive impairments.

Finally, there are differences in CNS lesions and their regional distribution. Lesions in HIV disease are found mostly in subcortical structures, including the basal ganglia, and some cortical nuclei (Price *et al*, 1988; Budka, 1991; Masliah *et al*, 1997; Stout *et al*, 1998; Everall *et al*, 1999). With MS, the principal lesion is demyelination—the destruction of oligodendrocyte processes forming the myelin sheath—although there is recent evidence of axonal transection too (Trapp *et al*, 1998). MS lesions are localized primarily in the optic nerve, spinal cord, brainstem, and cerebellum (McDonald and Ron, 1999). With AD, the pathological hallmarks are extracellular senile plaques and intraneuronal neurofibrillary tangles found in the cerebral cortex (temporal and parietal lobes), hippocampus and amygdala. Despite these differences, what becomes clear in subsequent sections are similarities in inflammatory processes and neuron dysfunction or death.

#### Chemokines and leukocyte chemotaxis

Chemokines are coming to prominence for their role as leukocyte chemoattractants in many CNS inflammatory diseases (Asensio and Campbell, 1999). Evidence of their role in regulating leukocyte infiltration across the BBB is being amassed for HIV (see prior discussion) and for MS (Ransohoff, 1999). Leukocyte infiltration across the BBB is not thought to be a major factor in AD because the inflammation is local, i.e.,

restricted within the brain (Xia and Hyman, 1999). Nevertheless, chemokines appear to play a more focal role in AD by attracting leukocytes residing *within* the CNS to senile plaques. In all three diseases, the accumulation of activated leukocytes within the CNS unleashes or contributes to inflammatory destruction of neurons or oligodendrocytes (see next section).

In MS, leukocyte infiltration is a major feature, apparently triggered by myelinreactive T cells which enter the CNS (Hickey *et al*, 1991; Brosnan and Raine, 1996; Ransohoff, 1999). The infiltrate consists mostly of macrophages and T cells (Cross *et al*, 1990, 1993), although B cells are increased later in disease (Ozawa *et al*, 1994). The chemokine profile in MS reveals upregulated expression of several  $\beta$ -chemokines (including RANTES and MCP-1) and IP-10, among others (Glabinski and Ransohoff, 1999). This chemokine profile resembles that with HIV, which, as noted before, consists of  $\beta$ -chemokines (MIP-1 $\alpha$ , MIP-1 $\beta$ , RANTES, MCP-3) and the  $\alpha$ -chemokine IP-10.

The significance of chemokines in MS pathogenesis has come from two types of evidence. First, antibodies against the chemokine MCP-1 block the onset of acute symptoms in an animal model of MS (Karpus *et al*, 1995). Second, studies of human CSF reveal upregulation of certain chemokines (and their cognate receptors) at the time of initial MS attack or relapse (Sorenson *et al*, 1999). The latter study is reminiscent of findings with HIV disease, cited earlier, of chemokine upregulation in CSF being correlated with the degree of dementia.

The dense senile plaque of AD consists of the core deposit of the protein  $\beta$ -amyloid surrounded by microglia, astrocytes, and dystrophic neurites (Selkoe, 1999). Responding to chemotactic signals from  $\beta$ -amyloid (Maeda *et al*, 1997; Kopec and Carroll, 1998), microglia arrive to digest  $\beta$ -amyloid, only to be incapable of degrading large amounts (Paresce *et al*, 1997). What signals draw microglia to  $\beta$ -amyloid deposits? A working hypothesis is that enhanced chemokine expression by astrocytes or microglia controls chemotaxis of other microglia to  $\beta$ -amyloid deposits (Xia *et al*, 1997; Xia and Hyman, 1999). A role for chemokines and their receptors in the neuropathogenesis of AD has been supported by finding that  $\beta$ -amyloid induces secretion of the chemokine IL-8 by cultured astrocytes (Gitter *et al*, 1995) and that neuritic portions of plaques express high levels of CXCR2, the chemokine receptor for IL-8 (Xia *et al*, 1997; Horuk *et al*, 1997).

#### *Activation of inflammatory cells and release of soluble cytotoxins*

A unifying theme in the neuropathogenesis of HIV, MS, and AD is the sustained overproduction and release of pro-inflammatory cytokines and other soluble factors by activated immune cells, resulting in eventual injury or death to nearby neurons and/or oligodendrocytes (Benveniste, 1998; Cotter *et al*,

1999; Gonzalez-Scarano and Baltuch, 1999; McGeer and McGeer, 1999).

Overproduction of toxic inflammatory factors applies across these three diseases, despite dissimilarities in (1) the specific sequence of events (which also may be heterogeneous at different stages of each disease and across different subtypes of each); (2) the relative contributions of different cytokines and soluble cytotoxins; (3) the affected region of the CNS and its proximity to vasculature; (4) the contributions of distinct classes of activated immunocompetent cells (T cells, infiltrating monocytes, resident microglia, and/or astrocytes); and (5) the nature and degree of injury and death to neurons (or oligodendrocytes). The *indirect* mechanisms of cytotoxicity outlined here do not preclude the co-occurrence of *direct* cytotoxicity mediated by cytotoxic T cells (CD8+) in MS (Brosnan and Raine, 1996) and by  $\beta$ -amyloid in AD (Yankner *et al*, 1990). This also does not preclude a role for B cells in antibody-mediated damage to oligodendrocytes in MS. Demyelination in animal models requires *both* autoreactive T cells and antibodies to myelin (Lassmann *et al*, 1988).

There are some interesting overlaps in secretory products between HIV, MS, and AD (except for gp120 and other HIV proteins). Common to all three diseases are elevated levels of cytokines IL-1, IL-6, TNF- $\alpha$ , and certain isoforms of TGF- $\beta$  (Benveniste, 1998). Similarly, all three have elevated levels of reactive oxygen intermediates, nitric oxide, and glutamate (released by inflammatory cells or neurons) (Piani *et al*, 1991; Bo *et al*, 1994; Stover *et al*, 1997; Lipton, 1998; Selkoe, 1999). Distinct immune cell types produce and secrete many of the same toxins. Low concentrations of these inflammatory products could be beneficial; however, high local concentrations of cytotoxins produced over a sustained period, regardless of cellular source, produce injury or death of neurons or oligodendrocytes. There is also recent evidence that dementia patients with HIV and AD both have greater percentages of monocytes in peripheral blood with the activation marker CD69 (Pulliam *et al*, 1997; Kusdra *et al*, 2000).

One of the best ways of assessing the significance of individual cytotoxins in disease is through correlation with disease progression. Levels of the cytokine TNF- $\alpha$  in human CSF are correlated with the degree of dementia with HIV (Wesselingh *et al*, 1993) and are correlated with relapse in MS (Sharief and Hentges, 1991). There has been no consistent evidence of cytokine levels in CSF increasing over the course of AD (Engelborghs *et al*, 1999; Lanzrein *et al*, 1998); however, there is new evidence of IL-1 overexpression correlating with dementia in AD (Nicoll *et al*, 2000). Establishing relationships between CSF levels and disease severity is complicated by the inherent limitations of using CSF levels to infer highly localized CNS inflammation. No single CSF marker for measuring progression has reached clinical use for the diseases discussed here.

### *Mechanisms of neuron dysfunction and death*

Refined understanding of the mechanisms of neurotoxicity can inspire targeted therapeutic strategies to reverse injury at the earliest possible stage or to prevent cell death. If the mechanisms are similar across different diseases, the same targeted treatments may be effective. The *in vivo* mechanisms of neuron dysfunction and death in HIV, MS, and AD are poorly understood, but there are numerous similarities to pursue.

At first glance the inclusion of MS might seem questionable, for MS largely features injury and demise of oligodendrocytes. Axons were thought to be protected, and clinical manifestations were thought to reflect conductance block by demyelinated, yet intact, axons. New research, however, has revealed axonal transection in MS (Trapp *et al*, 1998). Axonal loss is now postulated to be responsible for irreversible disability in MS (Trapp *et al*, 1998; Hickey, 1999b; Smith and McDonald, 1999). Axonal loss may be mediated by inflammatory factors in a manner similar to neurotoxicity with AD and HIV. After the death of oligodendrocytes, axons may become more vulnerable to the cytotoxic barrage from which they had previously been shielded by viable oligodendrocytes. High levels of some of the same cytokines are toxic to both neurons and oligodendrocytes (Selmaj and Raine, 1988; Benveniste, 1998). Although other neurotoxic mechanisms may be occurring (prolonged electrical silence, loss of trophic factors from oligodendrocytes, see review Scolding, 1999), neurotoxicity by inflammatory factors may be as salient a feature for MS as it appears to be for AD and HIV.

Much research has focused on neurotoxicity induced by excess levels of glutamate (Rothman and Olney, 1995; Choi, 1988). Excess levels of glutamate have been implicated, with varying levels of evidence, in all three diseases. In fact, excess glutamate has been implicated in so many neurological diseases that a landmark article referred to glutamate and other excitatory amino acids as a "final common pathway for neurologic disorders" (Lipton and Rosenberg, 1994). High extracellular levels of glutamate or glutamate agonists overstimulate N-methyl-D-aspartate (NMDA) receptors on neurons, resulting in the influx of calcium ions. High intracellular calcium levels, in turn, generate free radicals, activate proteases and phospholipase, and in-

duce mitochondrial dysfunction, leading to necrosis or apoptosis (Lipton, 1998; Martin *et al*, 1998). There are numerous pathways in which the cascade of intracellular events can unfold, for no single pathway of excitotoxicity is uniquely associated with cell death (Rothman and Olney, 1995; Klegeris and McGeer, 2000). Excess levels of glutamate are also toxic to oligodendrocytes, which possess the AMPA ( $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid)/kainate type of glutamate receptor (McDonald *et al*, 1998). Even if the intracellular mechanisms of cell death are less clear, the initial step may be the overactivation of glutamate receptors.

Research is beginning to explore the mechanisms of cytokine-induced neurotoxicity and its application to disease states. TNF- $\alpha$ , a cytokine implicated in HIV, MS, and AD, has apoptotic effects on neurons, possibly by reducing gene expression of Bcl-2, a protein that normally inhibits apoptosis (Pulliam *et al*, 1998). Research also has focused on IL-1, which appears to alter neuronal signaling by inhibiting synaptic transmission (Xiong *et al*, 2000). Signaling abnormalities are also beginning to be demonstrated after neuronal exposure to chemokines upregulated in disease, with effects mediated by various chemokine receptors (e.g. CXCR4) on neurons (Hesselgesser and Horuk, 1999; Zheng *et al*, 1999). This is a new and intensive area of investigation because chemokine receptors are expressed on neurons throughout the brain yet their normal functions are poorly understood. Through their activation of G-protein-coupled receptors, chemokine receptors may play diverse roles in synaptic transmission, signal transduction, and neuronal survival (Meucci *et al*, 1998).

### **Conclusions and treatment implications**

HIV neuropathogenesis research stands to benefit from better understanding of pathogenic relationships to systemic HIV disease and to other CNS diseases. These research directions will catalyze new avenues of treatment research. The classes of potential treatments include anti-inflammatories, NMDA blockers, chemokine receptor blockers, free radical scavengers, and antioxidants. The potential for cross-fertilization is driving a new era for HIV research.

### **References**

- Albert SM, Marder K, Dooneief G, Bell K, Sano M, Todak G, Stern Y (1995). Neuropsychologic impairment in early HIV infection. A risk factor for work disability. *Arch Neurol* **52**: 525–530.
- Asensio V, Campbell I (1999). Chemokines in the CNS: plurifunctional mediators in diverse states. *Trends Neurosci* **22**: 504–512.
- Aweeka F, Jayewardene A, Staprans S, Belibas SE, Kearney B, Lizak P, Novakovic-Agopian T, Price RW (1999). Failure to detect nelfinavir in the cerebrospinal fluid of HIV-1 infected patients with and without AIDS dementia complex. *J Acquir Immune Defic Syndr Hum Retrovirol* **20**: 39–43.
- Banks WA (1999). Physiology and pathology of the blood-brain barrier: Implications for microbial pathogenesis, drug delivery and neurodegenerative disorders. *J NeuroVirol* **5**: 538–555.

- Bell JE, Busuttill A, Ironside JE, Rebus S, Donaldson YK, Simmonds P, Peutherer JF (1993). Human immunodeficiency virus and the brain: Investigations of virus load and neuropathologic changes in pre-AIDS subjects. *J Infect Dis* **168**: 818–824.
- Benveniste E (1998). Cytokine actions in the central nervous system. *Cytokine Growth Factor Rev* **9**: 259–275.
- Bo L, Dawson T, Wesselingh S, Mork S, Choi S, Kong PA, Hanley D, Trapp BD (1994). Induction of nitric oxide synthase in demyelinating regions of multiple sclerosis brains. *Ann Neurol* **36**: 778–786.
- Brosnan C, Raine C (1996). Mechanisms of immune injury in multiple sclerosis. *Brain Pathol* **6**: 243–257.
- Budka H (1991). Neuropathology of human immunodeficiency virus infection. *Brain Pathol* **1**: 163–175.
- Center for Disease Control (CDC) (1999). New data show continued decline in AIDS deaths. <http://www.cdc.gov/od/oc/media/pressrel/r990831.htm>
- Choi DW (1988). Glutamate neurotoxicity and diseases of the nervous system. *Neuron* **1**: 623–634.
- Chun TW, Carruth L, Finzi D, Shen X, DiGiuseppe JA, Taylor H, Hemankova M, Chadwick K, Margolick J, Quinn TC, Kuo YH, Brookmeyer R, Zeiger MA, Barditch-Crovo P, Siliciano RF (1997). Quantification of latent tissue reservoirs and total body viral load in HIV-1 infection. *Nature* **387**: 183–188.
- Chun TW, Davey RT, Ostrowski M, Justement JS, Engel D, Mullins JI, Fauci AS (2000). Relationship between pre-existing viral reservoirs and the re-emergence of plasma viremia after discontinuation of highly active anti-retroviral therapy. *Nat Med* **6**: 757–761.
- Chun TW, Engel D, Mizell SB, Hallahan CW, Fischette M, Park S, Davey RT Jr, Dybul M, Kovacs JA, Metcalf JA, Mican JM, Berry MM, Corey L, Lane HC, Fauci AS (1999). Effect of interleukin-2 on the pool of latently infected, resting CD4+ T cells in HIV-1 infected patients receiving highly active anti-retroviral therapy. *Nat Med* **5**: 651–655.
- Chun TW, Fauci AS (1999). Latent reservoirs of HIV: Obstacles to the eradication of virus. *Proc Natl Acad Sci USA* **96**: 10958–10961.
- Cinque P, Vago L, Mengozzi M, Torri V, Ceresa D, Vicenzi E, Transidico P, Vagani A, Sozzani S, Mantovani A, Lazzarin A, Poli G (1998). Elevated cerebrospinal fluid levels of monocyte chemoattractant protein-1 correlated with HIV-1 encephalitis and local viral replication. *AIDS* **12**: 1327–1332.
- Conant K, Garzino-Demo A, Nath A, McArthur JC, Halliday W, Power C, Gallo RC, Major EO (1998). Induction of monocyte chemoattractant protein-1 in HIV-1 Tat-stimulated astrocytes and elevation in AIDS dementia. *Proc Natl Acad Sci USA* **95**: 3117–3121.
- Conlon P, Oksenberg JR, Zhang J, Steinman L (1999). The immunobiology of multiple sclerosis: an autoimmune disease of the central nervous system. *Neurobiol Dis* **6**: 149–166.
- Cotter RL, Burke WJ, Thomas VS, Potter JF, Zheng J, Gendelman HE (1999). Insights into the neurodegenerative process of Alzheimer's disease: a role for mononuclear phagocyte-associated inflammation and neurotoxicity. *J Leukoc Biol* **65**: 416–427.
- Cross AH, Cannella B, Brosnan CF, Raine CS (1990). Homing to central nervous system vasculature by antigen-specific lymphocytes. I. Localization of 14C-labeled cells during acute, chronic, and relapsing experimental allergic encephalomyelitis. *Lab Invest* **63**: 162–170.
- Cross AH, O'Mara T, Raine CS (1993). Chronic localization of myelin-reactive cells in the lesions of relapsing EAE: implications for the study of multiple sclerosis. *Neurology* **43**: 1028–1033.
- Dallasta LM, Pisarove LA, Esplen JE, Werley JV, Moses AV, Nelson JA, Achim CL (1999). Blood-brain barrier tight junction disruption in human immunodeficiency virus-1 encephalitis. *Am J Pathol* **155**: 1915–1927.
- Davis LE, Hjelle BL, Miller VE, Palmer DL, Llewellyn AL, Merlin TL, Young SA, Mills RG, Wachsman W, Wiley CA (1992). Early viral brain invasion in iatrogenic human immunodeficiency virus infection. *Neurology* **42**: 1736–1739.
- Dhawan S, Puri RK, Kumar A, Duplan H, Mason JM, Aggrawal BB (1997). Human immunodeficiency virus-1-tat protein induces the cell surface expression of endothelial leukocyte adhesion molecule-1, vascular cell adhesion molecule-1, and intercellular adhesion molecule-1 in human endothelial cells. *Blood* **90**: 1535–1544.
- Dickson DW, Lee SC, Mattiace LA, Yen SC, Brosnan C (1993). Microglia and cytokines in neurological disease, with special reference to AIDS and Alzheimer's disease. *Glia* **7**: 75–83.
- Donaldson YK, Bell JE, Ironside JW, Brettell RP, Roberston JR, Busuttill A, Simmonds P (1994). Redistribution of HIV outside the lymphoid system with onset of AIDS. *Lancet* **343**: 383–385.
- Dore GJ, Correll PK, Li Y, Kaldor JM, Cooper DA, Brew BJ (1999). Changes to AIDS dementia complex in the era of highly active antiretroviral therapy. *AIDS* **13**: 1249–1253.
- Ellis RJ, Deutsch R, Heaton RK, Marcotte TD, McCutchan JA, Nelson JA, Abramson I, Thal LJ, Atkinson JH, Wallace MR, Grant I (1997). Neurocognitive impairment is an independent risk factor for death in HIV infection. San Diego HIV Neurobehavioral Research Center Group. *Arch Neurol* **54**: 416–424.
- Ellis RJ, Gamst AC, Capparelli E, Spector SA, Hsia K, Wolfson T, Abramson I, Grant I, McCutchan JA (2000). Cerebrospinal fluid HIV RNA originates from both local CNS and systemic sources. *Neurology* **54**: 927–936.
- Engelborghs S, De Brabander M, De Cree J, D'Hooge R, Geerts H, Verhaegen H, De Deyn PP (1999). Unchanged levels of interleukins, neopterin, interferon-gamma and tumor necrosis factor-alpha in cerebrospinal fluid of patients with dementia of the Alzheimer type. *Neurochem Int* **34**: 523–530.
- Enting RH, Hoetelmans RM, Lange JM, Burger DM, Beijnen JH, Portegies P (1998). Antiretroviral drugs and the central nervous system. *AIDS* **12**: 1941–1955.
- Everall IP, Heaton RK, Marcotte TD, Ellis RJ, McCutchan JA, Atkinson JH, Grant I, Mallory M, Masliah E (1999). Cortical synaptic density is reduced in mild to moderate human immunodeficiency virus neurocognitive disorder. HNRC Group. HIV Neurobehavioral Research Center. *Brain Pathol* **9**: 209–217.
- Ferrando S, van Gorp W, McElhiney M, Goggin K, Sewell M, Rabkin J (1998). Highly active antiretroviral treatment in HIV infection: benefits for neuropsychological function. *AIDS* **12**: F65–F70.
- Finzi D, Blankson J, Siliciano JD, Margolick JB, Chadwick K, Pierson T, Smith K, Lisziewicz J, Lori F, Flexner C,



- Quinn TC, Chaisson RE, Rosenberg E, Walker B, Gange S, Gallant J, Siliciano RF (1999). Latent infection of CD4+ T cells provides a mechanism for lifelong persistence of HIV-1, even in patients on effective combination therapy. *Nat Med* **5**: 512–517.
- Finzi D, Hermankova M, Pierson T, Carruth LM, Buck C, Chaisson RE, Quinn TC, Chadwick K, Margolick J, Brookmeyer R, Gallant J, Markowitz M, Ho DD, Richman DD, Siliciano RF (1997). Identification of a reservoir for HIV-1 in patients on highly active antiretroviral therapy. *Science* **278**: 1295–1300.
- Flexner C (1998). HIV-protease inhibitors. *N Engl J Med* **338**: 1281–1292.
- Frankenberger M, Sternsdorf T, Pechumer H, Pforte A, Ziegler-Heitbrock HW (1996). Differential cytokine expression in human blood monocyte subpopulations: a polymerase chain reaction analysis. *Blood* **87**: 373–377.
- Gartner S (2000). HIV infection and dementia. *Science* **287**: 602–604.
- Gitter BD, Cox LM, Rydel RE, May PC (1995). Amyloid beta peptide potentiates cytokine secretion by interleukin-1 beta-activated human astrocytoma cells. *Proc Natl Acad Sci USA* **92**: 10738–10741.
- Giulian D, Vaca K, Noonan CA (1990). Secretion of neurotoxins by mononuclear phagocytes infected with HIV-1. *Science* **250**: 1593–1596.
- Glabinski AR, Ransohoff RM (1999). Sentries at the gate: chemokines and the blood-brain barrier. *J NeuroVirol* **5**: 623–634.
- Glass JD, Fedor H, Wesslingh SL, McArthur JC (1995). Immunocytochemical quantitation of human immunodeficiency virus in the brain: correlations with dementia. *Ann Neurol* **38**: 755–762.
- Gonzalez-Scarano F, Baltuch G (1999). Microglia as mediators of inflammatory and degenerative diseases. *Annu Rev Neurosci* **22**: 219–240.
- Grant I, Heaton RK, Atkinson JH (1995). Neurocognitive disorders in HIV-1 infection. HNRC Group. HIV Neurobehavioral Research Center. *Curr Top Microbiol Immunol* **202**: 11–32.
- Griffin DE (1998). HIV infection of the brain: viruses, cytokines, and immune regulatory factors associated with dementia. In: *The neurology of AIDS*. Gendelman HE, Lipton SA, Epstein L, Swindells, (eds). New York: Chapman and Hall, pp 73–85.
- Heaton RK, Marcotte TD, White DA, Ross D, Meredith K, Taylor MJ, Kaplan R, Grant I (1996). Nature and vocational significance of neuropsychiatric impairment associated with HIV infection. *Clin Neuropsychol* **10**: 1–14.
- Heaton RK, Velin RA, McCutchan JA, Gulevich SJ, Atkinson JH, Wallace MR, Godfrey HP, Kirson DA, Grant I (1994). Neuropsychological impairment in human immunodeficiency virus-infection: implications for employment. HNRC Group. HIV Neurobehavioral Research Center. *Psychosom Med* **56**: 8–17.
- Hesselgesser J, Horuk R (1999). Chemokine and chemokine receptor expression in the central nervous system. *J NeuroVirol* **5**: 13–26.
- Heyes MP, Jordan EK, Lee K, Saito K, Frank JA, Snoy PJ, Markey SP, Gravall M (1992). Relationship of neurologic status in macaques infected with the simian immunodeficiency virus to cerebral spinal fluid quinolinic acid and kynurenic acid. *Brain Res* **570**: 237–250.
- Hickey WF (1999a). Leukocyte traffic in the central nervous system: the participants and their roles. *Semin Immunol* **98**: 125–137.
- Hickey WF (1999b). The pathology of multiple sclerosis: a historical perspective. *J Neuroimmunol* **98**: 37–44.
- Hickey WF, Hsu BL, Kimura H (1991). T-lymphocyte entry into the central nervous system. *J Neurosci Res* **51**: 254–260.
- Hickey WF, Vass K, Lassmann H (1992). Bone marrow-derived elements in the central nervous system: an immunohistochemical and ultrastructural survey of rat chimeras. *J Neuropathol Exp Neurol* **51**: 246–256.
- Horuk R, Martin AW, Wang Z, Schweitzer L, Gerassimides A, Guo H, Lu Z, Hesselgesser J, Perez HD, Kim J, Parker J, Hadley TJ, Peiper SC (1997). Expression of chemokine receptors by subsets of neurons in the central nervous system. *J Immunol* **158**: 2882–2890.
- Hughes ES, Bell JE, Simmonds P (1997). Investigation of the dynamics of the spread of human immunodeficiency virus to brain and other tissues by evolutionary analysis of sequences from the p17gag and env genes. *J Virol* **71**: 1272–1280.
- Hurtrel B, Chakrabarti L, Hurtrel M, Maire MA, Dormont D, Montagnier L (1991). Early SIV encephalopathy. *J Med Primatol* **20**: 159–166.
- Imami N, Antonopoulos C, Hardy GA, Gazzard B, Gotch FM (1999). Assessment of type 1 and type 2 cytokines in HIV type 1-infected individuals: impact of highly active antiretroviral therapy. *AIDS Res Hum Retroviruses* **15**: 1499–1508.
- Karpus WJ, Lukacs NW, McRae BL, Strieter RM, Kunkel SL, Miller SD (1995). An important role for the chemokine macrophage inflammatory protein-1 alpha in the pathogenesis of the T cell-mediated autoimmune disease, experimental autoimmune encephalomyelitis. *J Immunol* **155**: 5003–5010.
- Kelder W, McArthur JC, Nance-Sproson T, McClernon D, Griffin DE (1998). Betachemokines MCP-1 and RANTES are selectively increased in cerebrospinal fluid of patients with human immunodeficiency virus-associated dementia. *Ann Neurol* **44**: 831–835.
- Kepler TB, Perelson AS (1998). Drug concentration heterogeneity facilitates the evolution of drug resistance. *Proc Natl Acad Sci USA* **95**: 11514–11519.
- Klegeris A, McGeer PL (2000). Interaction of various intracellular signaling mechanisms involved in mononuclear phagocyte toxicity toward neuronal cells. *J Leukoc Biol* **67**: 127–133.
- Kolb SA, Sporer B, Lahrtz F, Koedel U, Pfister HW, Fontana A (1999). Identification of a T cell chemotactic factor in the cerebrospinal fluid of HIV-1 infected individuals as interferon-gamma inducible protein 10. *J Neuroimmunol* **93**: 172–181.
- Kolson DL, Lavi E, Gonzalez-Scarano F (1998). The effects of human immunodeficiency virus in the central nervous system. *Adv Virus Res* **50**: 1–47.
- Kopec KK, Carroll RT (1998). Alzheimer's beta-amyloid peptide 1-42 induces a phagocytic response in murine microglia. *J Neurochem* **71**: 2123–2131.
- Korber BT, Kunstman KJ, Patterson BK, Furtado M, McEvelly MM, Levy R, Wolinsky SM (1994). Genetic differences between blood- and brain-derived viral sequences from human immunodeficiency virus type-1 infected patients: evidence of conserved

- elements in the V3 region of the envelope protein of brain-derived sequences. *J Virol* **68**: 7467–7481.
- Kusdra L, Rempel H, Yaffe K, Pulliam L (2000). Elevation of CD69+ monocyte/macrophages in patients with Alzheimer's disease. *Immunobiol* **202**: 26–33.
- Lackner AA, Smith MO, Munn RJ, Martfeld DJ, Gardner MB, Marx PA, Dandekar, S (1991). Localization of simian immunodeficiency virus in the central nervous system of rhesus monkeys. *Am J Pathol* **139**: 609–621.
- Lanzrein AS, Johnston CM, Perry VH, Jobst KA, King EM, Smith AD (1998). Longitudinal study of inflammatory factors in serum, cerebrospinal fluid, and brain tissue in Alzheimer disease: interleukin-1 beta, interleukin-6, interleukin-1 receptor antagonist, tumor necrosis factor-alpha, the soluble tumor necrosis factor receptors I and II, and alpha 1-antichymotrypsin. *Alzheimer Dis Assoc Disord* **12**: 215–227.
- Lassmann H (1999). The pathology of multiple sclerosis and its evolution. *Philos Trans R Soc Lond B Biol Sci* **354**: 1635–1640.
- Lassmann H, Brunner C, Bradl M, Linington C (1988). Experimental allergic encephalomyelitis: the balance between encephalitogenic T lymphocytes and demyelinating antibodies determines size and structure of demyelinated lesions. *Acta Neuropathol (Berl)* **75**: 566–576.
- Lassmann H, Schmied M, Vass K, Hickey WF (1993). Bone marrow derived elements and resident microglia in brain inflammation. *Glia* **7**: 19–24.
- Lipton SA (1998). Neuronal injury associated with HIV-1: approaches to treatment. *Annu Rev Pharmacol Toxicol* **38**: 159–177.
- Lipton SA, Rosenberg PA (1994). Excitatory amino acids as a final common pathway for neurologic disorders. *N Engl J Med* **330**: 613–622.
- Liu Y, Tang XP, McArthur JC, Scott J, Gartner S (2000). Analysis of human immunodeficiency virus type 1 gp 160 sequences from a patient with HIV dementia: evidence for monocyte trafficking into brain. *J Neuro Virol* **6 Suppl 1**: S70–81.
- Luster AD (1998). Chemokines—chemotactic cytokines that mediate inflammation. *N Engl J Med* **338**: 436–445.
- Maeda K, Nakai M, Maeda S, Kawamata T, Yamaguchi T, Tanaka C (1997). Possible different mechanisms between amyloid-beta (25–35)- and substance P-induced chemotaxis of murine microglia. *Gerontology* **43 Suppl 1**: 11–15.
- Martin C, Sonnerborg A, Svensson JO, Stahle L (1999). Indinavir-based treatment of HIV-1 infected patients: efficacy in the central nervous system. *AIDS* **13**: 1227–1232.
- Martin LJ, Al-Abdulla NA, Brambrink AM, Kirsch JR, Sieber FE, Portera-Cailliau C (1998). Neurodegeneration in excitotoxicity, global cerebral ischemia, and target deprivation: a perspective on the contributions of apoptosis and necrosis. *Brain Res Bull* **46**: 281–309.
- Masliah E, DeTeresa RM, Mallory ME, Hansen LA (2000). Changes in pathological findings at autopsy in AIDS cases for the last 15 years. *AIDS* **14**: 69–74.
- Masliah E, Heaton RK, Marcotte TD, Ellis RJ, Wiley CA, Mallory M, Achim CL, McCutchan JA, Nelson JA, Atkinson JH, Grant I (1997). Dendritic injury is a pathological substrate for human immunodeficiency virus-related cognitive disorders. HNRC Group. The HIV Neurobehavioral Research Center. *Ann Neurol* **42**: 963–972.
- McArthur JC, Hoover DR, Bacellar H, Miller EN, Cohen BA, Becker JT, Graham NMH, McArthur JH, Selnes OA, Jacobson LP, Visscher BR, Concha M, Saah A (1993). Dementia in AIDS patients: incidence and risk factors. *Neurology* **43**: 2245–2252.
- McDonald JW, Althomsons SP, Hyre KL, Choi DW, Goldberg MP (1998). Oligodendrocytes from forebrain are highly vulnerable to AMPA/kainate receptor-mediated excitotoxicity. *Nat Med* **4**: 291–297.
- McDonald WI, Ron MA (1999). Multiple sclerosis: the disease and its manifestations. *Philos Trans R Soc Lond B Biol Sci* **354**: 1615–1622.
- McGeer PL, McGeer EG (1999). Inflammation of the brain in Alzheimer's disease: implications for therapy. *J Leukoc Biol* **65**: 409–415.
- Meucci O, Fatatis A, Simen AA, Bushell TJ, Gray PW, Miller RJ (1998). Chemokines regulate hippocampal neuronal signaling and gp120 neurotoxicity. *Proc Natl Acad Sci USA* **95**: 14500–14505.
- Miller DW (1999). Immunobiology of the blood-brain barrier. *J Neuro Virol* **5**: 570–578.
- Montaner JS, Reiss P, Cooper D, Vella S, Harris M, Conway B, Wainberg MA, Smith D, Robinson P, Hall D, Myers M, Lange JM (1998). A randomized, double-blind trial comparing combinations of nevirapine, didanosine, and zidovudine for HIV-infected patients: the INCAS trial. Italy, The Netherlands, Canada, and Australia Study. *JAMA* **279**: 930–937.
- Morris A, Marsden M, Halcrow K, Hughes ES, Brett RP, Bell JE, Simmonds P (1999). Mosaic structure of the human immunodeficiency virus type 1 genome infecting lymphoid cells and the brain: evidence for frequent in vivo recombination events in the evolution of regional populations. *J Virol* **73**: 8720–8731.
- Mrak RE, Griffin WS (1997). The role of chronic self-propagating glial responses in neurodegeneration: implications for long-lived survivors of human immunodeficiency virus. *J Neuro Virol* **3**: 241–246.
- Navia BA, Cho ES, Petito CK, Price RW (1986b). The AIDS dementia complex: II. Neuropathology. *Ann Neurol* **19**: 525–535.
- Navia BA, Jordan BD, Price RW (1986a). The AIDS dementia complex: I. Clinical features. *Ann Neurol* **19**: 517–524.
- Nicoll JAR, Mrak RE, Graham DI, Stewart J, Wilcock G, MacGowan S, Esiri MM, Murray LS, Dewar D, Love S, Moss T, Griffin WSF (2000). Association of interleukin-1 gene polymorphisms with Alzheimer's disease. *Ann Neurol* **47**: 365–368.
- Nuovo GJ, Alfieri ML (1996). AIDS dementia is associated with massive, active HIV-1 infection and concomitant expression of several cytokines. *Mol Med* **2**: 358–366.
- Ozawa K, Suchanek G, Breitschopf H, Bruck W, Budka H, Jellinger K, Lassmann H (1994). Patterns of oligodendroglia pathology in multiple sclerosis. *Brain* **117(Pt 6)**: 1311–1322.
- Pang S, Vinters HV, Akashi T, O'Brien WA, Chen IS (1991). HIV-1 env sequence variation in brain tissue of patients with AIDS-related neurologic disease. *J Acquir Immun Defic Syndr* **4**: 1082–1092.
- Paresce DM, Chung H, Maxfield FR (1997). Slow degradation of aggregates of the Alzheimer's disease amyloid beta-protein by microglial cells. *J Biol Chem* **272**: 29390–29397.

- Persidsky Y, Stins M, Way D, Witte MH, Weinand M, Kim KS, Bock P, Gendelman HE, Fiala M (1997). A model for monocyte migration through the blood-brain barrier during HIV-1 encephalitis. *J Immunol* **158**: 3499–3510.
- Petito CK, Cash KS (1992). Blood-brain barrier abnormalities in the acquired immunodeficiency syndrome: immunohistochemical localization of serum proteins in postmortem brain. *Ann Neurol* **32**: 658–666.
- Petito CK, Chen H, Mastro AR, Torres-Munoz J, Roberts B, Wood C (1999). HIV infection of choroid plexus in AIDS and asymptomatic HIV-infected patients suggests that the choroid plexus may be a reservoir of productive infection. *J NeuroVirol* **5**: 670–677.
- Piani D, Frei K, Do KQ, Cuenod M, Fontana A (1991). Murine brain macrophages induced NMDA receptor-mediated neurotoxicity in vitro by secreting glutamate. *Neurosci Lett* **133**: 159–162.
- Powderly WG (2000). Current approaches to treatment for HIV-1 infection. *J NeuroVirol* **6 Suppl 1**: S8–S14.
- Price RW, Brew B, Sidtis J, Rosenblum M, Scheck A, Cleary P (1988). The brain in AIDS: central nervous system HIV-1 infection and AIDS dementia complex. *Science* **239**: 586–592.
- Price RW, Staprans S (1997). Measuring the “viral load” in cerebrospinal fluid in human immunodeficiency virus infection: window into brain infection? *Ann Neurol* **42**: 675–678.
- Pulliam L, Clarke JA, McGrath MS, Moore D, McGuire D.() Monokine products as predictors of AIDS dementia. *AIDS* **10**: 1495–1500.
- Pulliam LR, Gascon M, Stubblebine D, McGuire D, McGrath MS (1977). Unique monocyte subset in patients with AIDS dementia. *Lancet* **349**: 692–695.
- Pulliam L, Herndier BG, Tang NM, McGrath MS (1991). Human immunodeficiency virus-infected macrophages produce soluble factors that cause histological and neurochemical alterations in cultured human brains. *J Clin Invest* **87**: 503–512.
- Pulliam L, Zhou M, Stubblebine M, Bitler CM (1998). Differential modulation of cell death proteins in human brain cells by tumor necrosis factor alpha and platelet activating factor. *J Neurosci Res* **15**: 530–538.
- Ransohoff RM (1999). Mechanisms of inflammation in MS tissue: adhesion molecules and chemokines. *J Neuroimmunol* **98**: 57–68.
- Rausch DM, Heyes MP, Murray EA, Lendvay J, Sharer LR, Ward JM, Rehm S, Weihe E, Eiden LE (1994). Cytopathologic and neurochemical correlates of progression to motor/cognitive impairment in SIV-infected rhesus monkeys. *J Neuropathol Exp Neurol* **53**: 165–175.
- Rausch DM, Stover ES (2000). Neuroscience research in AIDS. *Prog Neuro-Psychopharmacol* in press.
- Richman DD (1996). HIV therapeutics. *Science* **272**: 1886–1888.
- Rothman SM, Olney JW (1995). Excitotoxicity and the NMDA receptor—still lethal after eight years. *Trends Neurosci* **18**: 57–58.
- Sacktor NC, Lyles RH, McFarlane G (1999b). HIV-1 related neurological disease incidence changes in the era of highly active antiretroviral therapy. *Neurology* **52**: A252–A253.
- Sacktor NC, Lyles RH, Skolasky RL, Anderson DE, McArthur JC, McFarlane G, Selnes OA, Becker JT, Cohen B, Wesch J, Miller EN (1999a). Combination antiretroviral therapy improves psychomotor speed performance in HIV-seropositive homosexual men. Multicenter AIDS Cohort Study (MACS). *Neurology* **52**: 1640–1647.
- Sacktor NC, Skolasky RL, Lyles RH, Esposito D, Selnes OA, McArthur JC (2000). Improvement in HIV-associated motor slowing after antiretroviral therapy including protease inhibitors. *J NeuroVirol* **6**: 84–88.
- Sanders VJ, Pittman CA, White MG, Wang G, Wiley CA, Achim CL (1998). Chemokines and receptors in HIV encephalitis. *AIDS* **12**: 1021–1026.
- Sasseville VG, Smith MM, Mackay CR, Pauley DR, Mansfield KG, Ringler DJ, Lackner AA (1996). Chemokine expression in simian immunodeficiency virus-induced AIDS encephalitis. *Am J Pathol* **149**: 1459–1467.
- Schmidtayerova H, Nottet HS, Nuovo G, Raabe T, Flanagan CR, Dubrovsky L, Gendelman HE, Cerami A, Bukrinsky M, Sherry B (1996). Human immunodeficiency virus type 1 infection alters chemokine beta peptide expression in human monocytes: implications for recruitment of leukocytes into brain and lymph nodes. *Proc Natl Acad Sci USA* **93**: 700–704.
- Schrager LK, D’Souza MP (1998). Cellular and anatomical reservoirs of HIV-1 in patients receiving potent antiretroviral combination therapy. *JAMA* **280**: 67–71.
- Scolding N (1999). Therapeutic strategies in multiple sclerosis. II. Long-term repair. *Philos Trans R Soc Lond B Biol Sci* **354**: 1711–1720.
- Sei S, Saito K, Stewart SK, Crowley JS, Brouwers P, Kleiner DE, Katz DA, Pizzo PA, Heyes MP (1995). Increased human immunodeficiency virus (HIV) type 1 DNA content and quinolinic acid concentration in brain tissues from patients with HIV encephalopathy. *J Infect Dis* **172**: 638–647.
- Selkoe DJ (1999). Translating cell biology into therapeutic advances in Alzheimer’s disease. *Nature* **399(6738 Suppl)**: A23–A31.
- Selmaj KW, Raine CS (1988). Tumor necrosis factor mediates myelin and oligodendrocyte damage in vitro. *Ann Neurol* **23**: 339–346.
- Shapshak P, Segal DM, Crandall KA, Fujimura RK, Zhang BT, Xin KQ, Okuda K, Petito CK, Eisdorfer C, Goodkin K (1999). Independent evolution of HIV type 1 in different brain regions. *AIDS Res Hum Retroviruses* **15**: 811–820.
- Sharer LR, Michaels J, Murphey-Corb M, Hu FS, Kueler DJ, Martin LN, Baskin GB (1991). Serial pathogenesis study of SIV brain infection. *J Med Primatol* **20**: 211–217.
- Sharief MK, Hentges R (1991). Association between tumor necrosis factor-alpha and disease progression in patients with multiple sclerosis. *N Engl J Med* **325**: 467–472.
- Sinclair E, Gray F, Ciardi A, Scaravilli F (1994). Immunohistochemical changes and PCR detection of HIV provirus DNA in brains of asymptomatic HIV-positive patients. *J Neuropathol Exp Neurol* **53**: 43–50.
- Smith KJ, McDonald WI (1999). The pathophysiology of multiple sclerosis: the mechanisms underlying the production of symptoms and the natural history of the disease. *Philos Trans R Soc Lond B Biol Sci* **354**: 1649–1673.
- Sorenson TL, Tani M, Jensen J, Pierce V, Lucchinetti C, Folchik VA, Quin S, Rottman J, Sellebjerg F, Stricter RM, Frederickson JL, Ransohoff RM (1999). Expression of specific chemokines and chemokine receptors in the central nervous system of multiple sclerosis patients. *J Clin Invest* **103**: 807–815.
- Stout JC, Ellis RJ, Jernigan TL, Archibald SL, Abramson I, Wolfson T, McCutchan JA, Wallace MR, Atkinson

- JH, Grant I (1998). Progressive cerebral volume loss in human immunodeficiency virus infection: a longitudinal volumetric magnetic resonance imaging study. HIV Neurobehavioral Research Center Group. *Arch Neurol* **55**: 161–168.
- Stover JF, Pleines UE, Morganti-Kossmann MC, Kossmann T, Lowitzsch K, Kempfski OS (1997). Neurotransmitters in cerebrospinal fluid reflect pathological activity. *Eur J Clin Invest* **27**: 1038–1043.
- Tozzi V, Balestra P, Galani S, Narciso P, Sebastiani G, D'Amato C, Affricano C, Pigorini F, Pau FM, De Felici A, Benedetto A (1999). Positive and sustained effects of highly active antiretroviral therapy on HIV-1 associated neurocognitive impairment. *AIDS* **13**: 1889–1997.
- Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mork S, Bo L (1998). Axonal transection in the lesions of multiple sclerosis. *N Engl J Med* **338**: 278–285.
- van't Wout AB, Ran LJ, Kuiken CL, Kookstra NA, Pals ST, Schuitemaker H (1998). Analysis of the temporal relationship between human immunodeficiency virus type 1 quasispecies in sequential blood samples and various organs obtained at autopsy. *J Virol* **72**: 488–496.
- Wesselingh SL, Power C, Glass JD, Tyor WR, McArthur JC, Farber JM, Griffin JW, Griffin DE (1993). Intracerebral cytokine messenger RNA expression in acquired immunodeficiency syndrome dementia. *Ann Neurol* **33**: 576–582.
- Westmoreland SV, Halpern E, Lackner AA (1998). Simian immunodeficiency virus encephalitis in rhesus macaques is associated with rapid disease progression. *J NeuroVirol* **4**: 260–268.
- Wiley CA, Achim CL, Christopherson C, Kidane Y, Kwok S, Masliah E, Mellors J, Radhakrishnan L, Wang G, Soontornniyomkij V (1999). HIV mediates a productive infection of the brain. *AIDS* **13**: 2055–2059.
- Wiley CA, Soontornniyomkij V, Radhakrishnan L, Masliah E, Mellors J, Hermann SA, Dailey P, Achim CL (1998). Distribution of brain HIV load in AIDS. *Brain Pathol* **8**: 277–284.
- Williams KC, Ulvestad E, Hickey WF (1994). Immunology of multiple sclerosis. *Clin Neurosci* **2**: 229–245.
- Wilt SG, Milward E, Zhou JM, Nagasato K, Patton H, Rusten R, Griffen DE, O'Connor M, Dubois-Dalcq M (1995). In vitro evidence for a dual role of tumor necrosis factor-alpha in human immunodeficiency virus type 1 encephalopathy. *Ann Neurol* **37**: 381–394.
- Wong JK, Hezareh M, Gunthard HF, Havlir DV, Ignacio CC, Spina CA, Richman DD (1997). Recovery of replication-competent HIV despite prolonged suppression of plasma viremia. *Science* **278**: 1291–1295.
- Wong JK, Ignacio CC, Torriani F, Havlir D, Fitch NJ, Richman DD (1997). In vivo compartmentalization of human immunodeficiency virus: evidence from the examination of pol sequences from autopsy tissues. *J Virol* **71**: 2059–2071.
- Xia M, Qin S, McNamara M, Mackay C, Hyman BT (1997). Interleukin-8 receptor B immunoreactivity in brain and neuritic plaques of Alzheimer's disease. *Am J Pathol* **150**: 1267–1274.
- Xia MQ, Hyman BT (1999). Chemokines/chemokine receptors in the central nervous system and Alzheimer's disease. *J NeuroVirol* **5**: 32–41.
- Xiong H, Zeng YC, Lewis T, Zheng J, Persidsky Y, Gendelman HE (2000). HIV-1 infected mononuclear phagocyte secretory products affect neuronal physiology leading to cellular demise: relevance for HIV-1 associated dementia. *J NeuroVirol* **6 Suppl 1**: S14–S23.
- Yankner BA, Duffy LK, Kirschner DA (1990). Neurotrophic and neurotoxic effects of amyloid beta protein: reversal by tachykinin neuropeptides. *Science* **250**: 279–282.
- Yeh MW, Kaul M, Zheng J, Nottet HS, Thylin M, Gendelman HE, Lipton SA (2000). Cytokine-stimulated, but not HIV-infected, human monocyte-derived macrophages produce neurotoxic levels of 1-cysteine. *J Immunol* **164**: 4265–4270.
- Zheng J, Gendelman HE (1997). The HIV-1 associated dementia complex: a metabolic encephalopathy fueled by viral replication in mononuclear phagocytes. *Curr Opin Neurol* **10**: 319–325.
- Zheng J, Thylin MR, Ghorpade A, Xiong H, Persidsky Y, Cotter R, Niemann D, Che M, Zeng YC, Gelbard HA, Shepard RB, Swartz JM, Gendelman HE (1999). Intracellular CXCR4 signaling, neuronal apoptosis and neuropathogenic mechanisms of HIV-1 associated dementia. *J Neuroimmunol* **98**: 185–200.
- Zink MC, Amedee AM, Mankowski JL, Craig L, Didier P, Carter DL, Munoz A, Murphey-Corb M, Clements JE (1997). Pathogenesis of SIV encephalitis. Selection and replication of neurovirulent SIV. *Am J Pathol* **151**: 793–803.
- Zink MC, Spelman JP, Robinson RB, Clements JE (1998). SIV infection of macaques—modeling the progression to AIDS dementia. *J NeuroVirol* **4**: 249–259.